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(54) Title: SECRETED AND TRANSMEMBRANE POLYPEPTIDES AND NUCLEIC ACIDS ENCODING THE SAME

(57) Abstract

The present invention is directed to novel polypeptides and to nucleic acid molecules encoding those polypeptides. Also provided herein are vectors and host cells comprising those nucleic acid sequences, chimeric polypeptides molecules comprising the polypeptides of the present invention fused to heterologous polypeptide sequences, antibodies which bind to the polypeptides of the present invention and to methods for producing the polypeptides of the present invention.

SECRETED AND TRANSMEMBRANE POLYPEPTIDES AND NUCLEIC ACIDS ENCODING THE SAME

FIELD OF THE INVENTION

The present invention relates generally to the identification and isolation of novel DNA and to the recombinant production of novel polypeptides encoded by that DNA.

BACKGROUND OF THE INVENTION

Extracellular and membrane-bound proteins play important roles in the formation, differentiation and maintenance of multicellular organisms. The fate of many individual cells, e.g., proliferation, migration, differentiation, or interaction with other cells, is typically governed by information received from other cells and/or the immediate environment. This information is often transmitted by secreted polypeptides (for instance, mitogenic factors, survival factors, cytotoxic factors, differentiation factors, neuropeptides, and hormones) which are, in turn, received and interpreted by diverse cell receptors or membrane-bound proteins. These secreted polypeptides or signaling molecules normally pass through the cellular secretory pathway to reach their site of action in the extracellular environment, usually at a membrane-bound receptor protein.

Secreted proteins have various industrial applications, including use as pharmaceuticals, diagnostics, biosensors and bioreactors. In fact, most protein drugs available at present, such as thrombolytic agents, interferons, interleukins, erythropoietins, colony stimulating factors, and various other cytokines, are secretory proteins. Their receptors, which are membrane-bound proteins, also have potential as therapeutic or diagnostic agents. Receptor immunoadhesins, for instance, can be employed as therapeutic agents to block receptor-ligand interaction. Membrane-bound proteins can also be employed for screening of potential peptide or small molecule inhibitors of the relevant receptor/ligand interaction. Such membrane-bound proteins and cell receptors include, but are not limited to, cytokine receptors, receptor kinases, receptor phosphatases, receptors involved in cell-cell interactions, and cellular adhesin molecules like selectins and integrins. Transduction of signals that regulate cell growth and differentiation is regulated in part by phosphorylation of various cellular proteins. Protein tyrosine kinases, enzymes that catalyze that process, can also act as growth factor receptors. Examples include fibroblast growth factor receptor and nerve growth factor receptor.

Efforts are being undertaken by both industry and academia to identify new, native secreted and membrane-bound receptor proteins. Many efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel secreted and membrane-bound receptor proteins. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)].

We herein describe the identification and characterization of novel secreted and transmembrane polypeptides and novel nucleic acids encoding those polypeptides.

and most blood cells. CR1 interacts with complement proteins C3b, C4b, and iC3b to accelerate dissociation of C3 convertases, acts as a cofactor for Factor I-mediated cleavage of C3b and C4b, and binds immune complexes and promotes their dissolution and phagocytosis.

Proteins which have homology to complement proteins are of particular interest to the medical and industrial communities. Often, proteins having homology to each other have similar function. It is also of interest when proteins having homology do not have similar functions, indicating that certain structural motifs identify information other than function, such as locality of function.

Efforts are being undertaken by both industry and academia to identify new, native secreted and membrane-bound proteins, particularly those having homology to known proteins involved in the complement pathway. Proteins involved in the complement pathway were reviewed in Birmingham DJ (1995), Critical Reviews in Immunology, 15(2):133-154 and in Abbas AK, et al. (1994) Cellular and Molecular Immunology, 2nd Ed. W.B. Saunders Company, Philadelphia, pp 295-315.

We herein describe the identification and characterization of novel polypeptides having homology to complement receptors, designated herein as PRO222 polypeptides.

21. PRO234

The successful function of many systems within multicellular organisms is dependent on cell-cell interactions. Such interactions are affected by the alignment of particular ligands with particular receptors in a manner which allows for ligand-receptor binding and thus a cell-cell adhesion. While protein-protein interactions in cell recognition have been recognized for some time, only recently has the role of carbohydrates in physiologically relevant recognition been widely considered (see B.K. Brandley *et al.*, *J. Leuk. Biol.* 40: 97 (1986) and N. Sharon *et al.*, *Science* 246: 227 (1989). Oligosaccharides are well positioned to act as recognition novel lectins due to their cell surface location and structural diversity. Many oligosaccharide structures can be created through the differential activities of a smaller number of glycosyltransferases. The diverse structures of oligosaccharides can be generated by transcription of relatively few gene products, which suggests that the oligosaccharides are a plausible mechanism by which is directed a wide range of cell-cell interactions. Examples of differential expression of cell surface carbohydrates and putative carbohydrate binding proteins (lectins) on interacting cells have been described (J. Dodd & T.M. Jessel, *J. Neurosci.* 5: 3278 (1985); L.J. Regan *et al.*, *Proc. Natl. Acad. Sci. USA* 83: 2248 (1986); M. Constantine-Paton *et al.*, *Nature* 324: 459 (1986); and M. Tiemeyer *et al.*, *J. Biol. Chem.* 263: 1671 (1989). One interesting member of the lectin family are selectins.

The migration of leukocytes to sites of acute or chronic inflammation involves adhesive interactions between these cells and the endothelium. This specific adhesion is the initial event in the cascade that is initiated by inflammatory insults, and it is, therefore, of paramount importance to the regulated defense of the organism.

The types of cell adhesion molecules that are involved in the interaction between leukocytes and the endothelium during an inflammatory response currently stands at four: (1) selectins; (2) (carbohydrate and glycoprotein) ligands for selectins; (3) integrins; and (4) integrin ligands, which are members of the immunoglobulin gene superfamily.

The selectins are cell adhesion molecules that are unified both structurally and functionally. Structurally, selectins are characterized by the inclusion of a domain with homology to a calcium-dependent lectin (C-lectins), an epidermal growth factor (egf)-like domain and several complement binding-like domains, Bevilacqua, M.P. *et al.*, *Science* 243: 1160-1165 (1989); Johnston *et al.*, *Cell* 56: 1033-1044 (1989); Lasky *et al. Cell* 56: 1045-1055 (1989); Siegalman, M. *et al.*, *Science* 243: 1165-1172 (1989); Stoolman, L.M., *Cell* 56: 907-910 (1989). Functionally, selectins share the common property of their ability to mediate cell binding through interactions between their lectin domains and cell surface carbohydrate ligands (Brandley, B, *et al.*, *Cell* 63, 861-863 (1990); Springer, T. and Lasky, L.A., *Nature* 349, 19-197 (1991); Bevilacqua, M.P. and Nelson, R.M., *J. Clin. Invest.* 91 379-387 (1993) and Tedder *et al.*, *J. Exp. Med.* 170: 123-133 (1989).

There are three members identified so far in the selectin family of cell adhesion molecules: L-selectin (also called peripheral lymph node homing receptor (pnHR), LEC-CAM-1, LAM-1, gp90^{MEL}, gp100^{MEL}, gp110^{MEL}, MEL-14 antigen, Leu-8 antigen, TQ-1 antigen, DREG antigen), E-selectin (LEC-CAM-2, LECAM-2, ELAM-1) and P-selectin (LEC-CAM-3, LECAM-3, GMP-140, PADGEM).

The identification of the C-lectin domain has led to an intense effort to define carbohydrate binding ligands for proteins containing such domains. E-selectin is believed to recognize the carbohydrate sequence NeuNAcα2-3Galβ1-4(Fucα1-3)GlcNAc (sialyl-Lewis x, or sLe^x) and related oligosaccharides, Berg *et al.*, *J. Biol. Chem.* 265: 14869-14872 (1991); Lowe *et al.*, *Cell* 63: 475-484 (1990); Phillips *et al.*, *Science* 250: 1130-1132 (1990); *Tiemeyer et al., Proc. Natl. Acad. Sci. USA* 88: 1138-1142 (1991).

L-selectin, which comprises a lectin domain, performs its adhesive function by recognizing carbohydrate-containing ligands on endothelial cells. L-selectin is expressed on the surface of leukocytes, such as lymphocytes, neutrophils, monocytes and eosinophils, and is involved with the trafficking of lymphocytes to peripheral lymphoid tissues (*Gallatin et al., Nature* 303: 30-34 (1983)) and with acute neutrophil-medicated inflammatory responses (Watson, S.R., *Nature* 349: 164-167 (1991)). The amino acid sequence of L-selectin and the encoding nucleic acid sequence are, for example, disclosed in U.S. patent No. 5,098,833 issued 24 March 1992.

L-selectin (LECAM-1) is particularly interesting because of its ability to block neutrophil influx (Watson *et al., Nature* 349: 164-167 (1991). It is expressed in chronic lymphocytic leukemia cells which bind to HEV (Spertini *et al., Nature* 349: 691-694 (1991). It is also believed that HEV structures at sites of chronic inflammation are associated with the symptoms of diseases such as rheumatoid arthritis, psoriasis and multiple sclerosis.

E-selectin (ELAM-1), is particularly interesting because of its transient expression on endothelial cells in response to IL-1 or TNF. Bevilacqua et al., *Science* 243: 1160 (1989). The time course of this induced expression (2-8 h) suggests a role for this receptor in initial neutrophil induced extravasation in response to infection and injury. It has further been reported that anti-ELAM-1 antibody blocks the influx of neutrophils in a primate asthma model and thus is beneficial for preventing airway obstruction resulting from the inflammatory response. Gundel *et al., J. Clin. Invest.* 88: 1407 (1991).

The adhesion of circulating neutrophils to stimulated vascular endothelium is a primary event of the inflammatory response. P-selectin has been reported to recognize the Lewis x structure (Galβ1-4(Fucα1-3) GlcNAc), Larsen *et al., Cell* 63: 467-474(1990). Others report that an additional terminal linked sialic acid is required for high affinity binding, Moore *et al., J. Cell. Biol.* 112: 491-499 (1991). P-selectin has been shown to be significant in acute

lung injury. Anti-P-selectin antibody has been shown to have strong protective effects in a rodent lung injury model. M.S. Mulligan *et al.*, *J. Clin. Invest.* 90: 1600 (1991).

We herein describe the identification and characterization of novel polypeptides having homology to lectin proteins, herein designated as PRO234 polypeptides.

22. PRO231

Some of the most important proteins involved in the above described regulation and modulation of cellular processes are the enzymes which regulate levels of protein phosphorylation in the cell. For example, it is known that the transduction of signals that regulate cell growth and differentiation is regulated at least in part by phosphorylation and dephosphorylation of various cellular proteins. The enzymes that catalyze these processes include the protein kinases, which function to phosphorylate various cellular proteins, and the protein phosphatases, which function to remove phosphate residues from various cellular proteins. The balance of the level of protein phosphorylation in the cell is thus mediated by the relative activities of these two types of enzymes.

Protein phosphatases represent a growing family of enzymes that are found in many diverse forms, including both membrane-bound and soluble forms. While many protein phosphatases have been described, the functions of only a very few are beginning to be understood (Tonks, *Semin. Cell Biol.* 4:373-453 (1993) and Dixon, *Recent Prog. Horm. Res.* 51:405-414 (1996)). However, in general, it appears that many of the protein phosphatases function to modulate the positive or negative signals induced by various protein kinases. Therefore, it is likely that protein phosphatases play critical roles in numerous and diverse cellular processes.

Given the physiological importance of the protein phosphatases, efforts are being undertaken by both industry and academia to identify new, native phosphatase proteins. Many of these efforts are focused on the screening of mammalian recombinant DNA libraries to identify the coding sequences for novel phosphatase proteins. Examples of screening methods and techniques are described in the literature [see, for example, Klein et al., Proc. Natl. Acad. Sci., 93:7108-7113 (1996); U.S. Patent No. 5,536,637)].

We herein describe the identification and characterization of novel polypeptides having homology to acid phosphatases, designated herein as PRO231 polypeptides.

23. PRO229

Scavenger receptors are known to protect IgG molecules from catabolic degradation. Riechmann and Hollinger, Nature Biotechnology, 15:617 (1997). In particular, studies of the CH2 and CH3 domains have shown that specific sequences of these domains are important in determining the half-lives of antibodies. Ellerson, et al., J. Immunol., 116: 510 (1976); Yasmeen, et al., J. Immunol. 116: 518 (1976; Pollock, et al., Eur. J. Immunol., 20: 2021 (1990). Scavenger receptor proteins and antibodies thereto are further reported in U.S. Patent No. 5,510,466 to Krieger, et al. Due to the ability of scavenger receptors to increase the half-life of polypeptides and their involvement in immune function, molecules having homology to scavenger receptors are of importance to the scientific and medical community.

Efforts are being undertaken by both industry and academia to identify new, native secreted and membrane-bound receptor proteins, particularly those having homology to scavenger receptors. Many efforts are focused on

polypeptide encoded by the nucleic acid deposited under accession number ATCC 209432.

19. PRO224

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO224".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO224 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO224 polypeptide having amino acid residues 1 to 282 of Figure 46 (SEQ ID NO:127), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO224 polypeptide. In particular, the invention provides isolated native sequence PRO224 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 282 of Figure 46 (SEQ ID NO:127).

20. PRO222

Applicants have identified a cDNA clone that encodes a novel polypeptide, wherein the polypeptide is designated in the present application as "PRO222".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO222 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO222 polypeptide having amino acid residues 1 to 490 of Fig. 48 (SEQ ID NO:132), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO222 polypeptide. In particular, the invention provides isolated native sequence PRO222 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 490 of Figure 48 (SEQ ID NO:132).

21. PRO234

Applicants have identified a cDNA clone that encodes a novel lectin polypeptide molecule, designated in the present application as "PRO234".

In one embodiment, the invention provides an isolated nucleic acid encoding a novel lectin comprising DNA encoding a PRO234 polypeptide. In one aspect, the isolated nucleic acid comprises the DNA encoding PRO234 polypeptides having amino acid residues 1 to 382 of Fig. 50 (SEQ ID NO:137), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions. In another aspect, the invention provides an isolated nucleic acid molecule comprising the nucleotide sequence of Fig. 49 (SEQ ID NO:136).

In another embodiment, the invention provides isolated novel PRO234 polypeptides. In particular, the invention provides isolated native sequence PRO234 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 382 of Figure 50 (SEQ ID NO:137).

In yet another embodiment, the invention provides oligonucleotide probes useful for isolating genomic and cDNA nucleotide sequences.

22. PRO231

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to a putative acid phosphatase, wherein the polypeptide is designated in the present application as "PRO231".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO231 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO231 polypeptide having amino acid residues 1 to 428 of Fig. 52 (SEQ ID NO:142), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO231 polypeptide. In particular, the invention provides isolated native sequence PRO231 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 428 of Figure 52 (SEQ ID NO:142).

23. PRO229

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to scavenger receptors wherein the polypeptide is designated in the present application as "PRO229".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO229 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO229 polypeptide having amino acid residues 1 to 347 of Figure 54 (SEQ ID NO:148), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO229 polypeptide. In particular, the invention provides isolated native sequence PRO229 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 347 of Figure 54 (SEQ ID NO:148).

24. PRO238

Applicants have identified a cDNA clone that encodes a novel polypeptide having homology to reductase, wherein the polypeptide is designated in the present application as "PRO238".

In one embodiment, the invention provides an isolated nucleic acid molecule comprising DNA encoding a PRO238 polypeptide. In one aspect, the isolated nucleic acid comprises DNA encoding the PRO238 polypeptide having amino acid residues 1 to 310 of Figure 56 (SEQ ID NO:153), or is complementary to such encoding nucleic acid sequence, and remains stably bound to it under at least moderate, and optionally, under high stringency conditions.

In another embodiment, the invention provides isolated PRO238 polypeptide. In particular, the invention provides isolated native sequence PRO238 polypeptide, which in one embodiment, includes an amino acid sequence comprising residues 1 to 310 of Figure 56 (SEQ ID NO:153).

Figure 46 shows the amino acid sequence (SEQ ID NO:127) derived from the coding sequence of SEQ ID NO:126 shown in Figure 45.

Figure 47 shows a nucleotide sequence (SEQ ID NO:131) of a native sequence PRO222 cDNA, wherein SEQ ID NO:131 is a clone designated herein as "UNQ196" and/or "DNA33107-1135".

Figure 48 shows the amino acid sequence (SEQ ID NO:132) derived from the coding sequence of SEQ ID NO:131 shown in Figure 47.

Figure 49 shows a nucleotide sequence (SEQ ID NO:136) of a native sequence PRO234 cDNA, wherein SEQ ID NO:136 is a clone designated herein as "UNQ208" and/or "DNA35557-1137".

Figure 50 shows the amino acid sequence (SEQ ID NO:137) derived from the coding sequence of SEQ ID NO:136 shown in Figure 49.

Figure 51 shows a nucleotide sequence (SEQ ID NO:141) of a native sequence PRO231 cDNA, wherein SEQ ID NO:141 is a clone designated herein as "UNQ205" and/or "DNA34434-1139".

Figure 52 shows the amino acid sequence (SEQ ID NO:142) derived from the coding sequence of SEQ ID NO:141 shown in Figure 51.

Figure 53 shows a nucleotide sequence (SEQ ID NO:147) of a native sequence PRO229 cDNA, wherein SEQ ID NO:147 is a clone designated herein as "UNQ203" and/or "DNA33100-1159".

Figure 54 shows the amino acid sequence (SEQ ID NO:148) derived from the coding sequence of SEQ ID NO:147 shown in Figure 53.

Figure 55 shows a nucleotide sequence (SEQ ID NO:152) of a native sequence PRO238 cDNA, wherein SEQ ID NO:152 is a clone designated herein as "UNQ212" and/or "DNA35600-1162".

Figure 56 shows the amino acid sequence (SEQ ID NO:153) derived from the coding sequence of SEQ ID NO:152 shown in Figure 55.

Figure 57 shows a nucleotide sequence (SEQ ID NO:158) of a native sequence PRO233 cDNA, wherein SEQ ID NO:158 is a clone designated herein as "UNQ207" and/or "DNA34436-1238".

Figure 58 shows the amino acid sequence (SEQ ID NO:159) derived from the coding sequence of SEQ ID NO:158 shown in Figure 57.

Figure 59 shows a nucleotide sequence (SEQ ID NO:163) of a native sequence PRO223 cDNA, wherein SEQ ID NO:163 is a clone designated herein as "UNQ197" and/or "DNA33206-1165".

Figure 60 shows the amino acid sequence (SEQ ID NO:164) derived from the coding sequence of SEQ ID NO:163 shown in Figure 59.

Figure 61 shows a nucleotide sequence (SEQ ID NO:169) of a native sequence PRO235 cDNA, wherein SEQ ID NO:169 is a clone designated herein as "UNQ209" and/or "DNA35558-1167".

Figure 62 shows the amino acid sequence (SEQ ID NO:170) derived from the coding sequence of SEQ ID NO:169 shown in Figure 61.

Figure 63 shows a nucleotide sequence (SEQ ID NO:174) of a native sequence PRO236 cDNA, wherein SEQ ID NO:174 is a clone designated herein as "UNQ210" and/or "DNA35599-1168".

Figure 64 shows the amino acid sequence (SEQ ID NO:175) derived from the coding sequence of SEQ ID NO:174 shown in Figure 63.

comprising amino acids 1 to 312 of Figure 24 (SEQ ID NO:64), the native sequence of each PRO220, PRO221 and PRO227 polypeptides is a mature or full-length native sequence PRO220, PRO221 and PRO227 polypeptide comprising amino acids 1 through 708 of Figure 26 (SEQ ID NO:69), 1 through 259 of Figure 28 (SEQ ID NO:71), and 1 through 620 of Figure 30 (SEQ ID NO:73), the native sequence PRO258 polypeptide is a mature or full-length native sequence PRO258 polypeptide comprising amino acids 1 to 398 of Figure 32 (SEQ ID NO:84) or the native sequence PRO258 polypeptide is an extracellular domain of the full-length PRO258 protein, wherein the putative transmembrane domain of the full-length PRO258 protein is encoded by nucleotides beginning at nucleotide 1134 of SEQ ID NO:83, the native sequence PRO266 polypeptide is a mature or full-length native sequence PRO266 polypeptide comprising amino acids 1 to 696 of Figure 34 (SEQ ID NO:91) or the native sequence PRO266 polypeptide is an extracellular domain of the full-length PRO266 protein, wherein the putative transmembrane domain of the full-length PRO266 protein is encoded by nucleotides beginning at about nucleotide 2009 of SEQ ID NO:104, the native sequence PRO269 polypeptide is a mature or full-length native sequence PRO269 polypeptide comprising amino acids 1 to 490 of Figure 36 (SEQ ID NO:96) or the native sequence PRO269 polypeptide is an extracellular domain of the full-length PRO269 protein, wherein the putative transmembrane domain of the full-length PRO269 protein is encoded by nucleotides beginning at nucleotide 1502 as shown in Figure 35, the native sequence PRO287 polypeptide is a mature or full-length native sequence PRO287 polypeptide comprising amino acids 1 to 415 of Figure 38 (SEQ ID NO:104), the native sequence PRO214 is a mature or full-length native sequence PRO214 comprising amino acids 1 to 420 of Fig. 40 (SEQ ID NO:109), the native-sequence PRO317 is a full-length native-pre-sequence PRO317 comprising amino acids 1 to 366 of Fig. 42 (SEQ ID NO:114) or a mature native-sequence PRO317 comprising amino acids 19 to 366 of Fig. 42 (SEQ ID NO:114), the native sequence PRO301 is a mature or full-length native sequence PRO301 comprising amino acids 1 to 299 of Fig. 44 (SEQ ID NO:119), with or without the N-terminal signal sequence, with or without the initiating methionine at position 1, with or without the potential transmembrane domain at position 236 to about 258, and with or without the intracellular domain at about position 259 to 299, the native sequence PRO224 polypeptide is a mature or full-length native sequence PRO224 polypeptide comprising amino acids 1 to 282 of Figure 46 (SEQ ID NO:127), the native sequence PRO222 polypeptide is a mature or full-length native sequence PRO222 polypeptide comprising amino acids 1 to 490 of Figure 48 (SEQ ID NO:132), the native sequence PRO234 is a mature or full-length native sequence novel lectin comprising amino acids 1 to 382 of Fig. 50 (SEQ ID NO:137), the native sequence PRO231 polypeptide is a mature or full-length native sequence PRO231 polypeptide comprising amino acids 1 to 428 of Figure 52 (SEQ ID NO:142), the native sequence PRO229 polypeptide is a mature or full-length native sequence PRO229 polypeptide comprising amino acids 1 to 347 of Figure 54 (SEQ ID NO:148), the native sequence PRO238 polypeptide is a mature or full-length native sequence PRO238 polypeptide comprising amino acids 1 to 310 of Figure 56 (SEQ ID NO:153), the native sequence PRO233 polypeptide is a mature or full-length native sequence PRO233 polypeptide comprising amino acids 1 to 300 of Figure 58 (SEQ ID NO:159), the native sequence PRO223 polypeptide is a mature or full-length native sequence PRO223 polypeptide comprising amino acids 1 to 476 of Figure 60 (SEQ ID NO:164), the native sequence PRO235 polypeptide is a mature or full-length native sequence PRO235 polypeptide comprising amino acids 1 to 552 of Figure 62 (SEQ ID NO:170), the native sequence PRO236 polypeptide is a mature or full-length native sequence PRO236 polypeptide comprising amino acids 1 to 636 of Figure 64 (SEQ ID NO:175), the native sequence PRO262

complement C3b receptor type 2 long form precursor, has 40% amino acid identity with human hypothetical protein kiaa0247. Accordingly, it is presently believed that PRO222 polypeptide disclosed in the present application is a newly identified member of the complement receptor family and possesses activity typical of the complement receptor family.

21. Full-length PRO234 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO234. In particular, Applicants have identified and isolated cDNA encoding a PRO234 polypeptide, as disclosed in further detail in the Examples below. Using BLAST (FastA-format) sequence alignment computer programs, Applicants found that a cDNA sequence encoding full-length native sequence PRO234 has 31% identity and Blast score of 134 with E-selectin precursor. Accordingly, it is presently believed that the PRO234 polypeptides disclosed in the present application are newly identified members of the lectin/selectin family and possess activity typical of the lectin/selectin family.

22. Full-length PRO231 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO231. In particular, Applicants have identified and isolated cDNA encoding a PRO231 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that the full-length native sequence PRO231 polypeptide (shown in Figure 52 and SEQ ID NO:142) has 30 % and 31 % amino acid identity with human and rat prostatic acid phosphatase precursor proteins, respectively. Accordingly, it is presently believed that the PRO231 polypeptide disclosed in the present application may be a newly identified member of the acid phosphatase protein family.

23. Full-length PRO229 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO229. In particular, Applicants have identified and isolated cDNA encoding a PRO229 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA sequence alignment computer programs, Applicants found that various portions of the PRO229 polypeptide have significant homology with antigen wc1.1, M130 antigen, T cell surface glycoprotein CD6 and CD6. It also is related to Sp-alpha. Accordingly, it is presently believed that PRO229 polypeptide disclosed in the present application is a newly identified member of the family containing scavenger receptor homology, a sequence motif found in a number of proteins involved in immune function and thus possesses immune function and /or segments which resist degradation, typical of this family.

24. Full-length PRO238 Polypeptides

The present invention provides newly identified and isolated nucleotide sequences encoding polypeptides referred to in the present application as PRO238. In particular, Applicants have identified and isolated cDNA encoding a PRO238 polypeptide, as disclosed in further detail in the Examples below. Using BLAST and FastA

of PLGA, lactic and glycolic acids, can be cleared quickly within the human body. Moreover, the degradability of this polymer can be adjusted from months to years depending on its molecular weight and composition. Lewis, "Controlled release of bioactive agents from lactide/glycolide polymer," in: M. Chasin and R. Langer (Eds.), Biodegradable Polymers as Drug Delivery Systems (Marcel Dekker: New York, 1990), pp. 1-41.

For example, for a formulation that can provide a dosing of approximately 80 g/kg/day in mammals with a maximum body weight of 85 kg, the largest dosing would be approximately 6.8 mg PRO317 per day. In order to achieve this dosing level, a sustained- release formulation which contains a maximum possible protein loading (15-20% w/w PRO317) with the lowest possible initial burst (<20%) is necessary. A continuous (zero-order) release of PRO317 from microparticles for 1-2 weeks is also desirable. In addition, the encapsulated protein to be released should maintain its integrity and stability over the desired release period.

It is contemplated that conditions or diseases of the uterus, endometrial tissue, or other genital tissues or cardiac tissues may precipitate damage that is treatable with PRO317 or PRO317 agonist where PRO317 expression is reduced in the diseased state; or with antibodies to PRO317 or other PRO317 antagonists where the expression of PRO317 is increased in the diseased state. These conditions or diseases may be specifically diagnosed by the probing tests discussed above for physiologic and pathologic problems which affect the function of the organ.

The PRO317, PRO317 agonist, or PRO317 antagonist may be administered to a mammal with another biologically active agent, either separately or in the same formulation to treat a common indication for which they are appropriate. For example, it is contemplated that PRO317 can be administered together with EBAF-1 for those indications on which they demonstrate the same qualitative biological effects. Alternatively, where they have opposite effects, EBAF-1 may be administered together with an antagonist to PRO317, such as an anti-PRO317 antibody. Further, PRO317 may be administered together with VEGF for coronary ischemia where such indication is warranted, or with an anti-VEGF for cancer as warranted, or, conversely, an antagonist to PRO317 may be administered with VEGF for coronary ischemia or with anti-VEGF to treat cancer as warranted. These administrations would be in effective amounts for treating such disorders.

Native PRO301 (SEQ ID NO:119) has a Blast score of 246 and 30% homology at residues 24 to 282 of Figure 44 with A33_HUMAN, an A33 antigen precursor. A33 antigen precursor, as explained in the Background is a tumor-specific antigen, and as such, is a recognized marker and therapeutic target for the diagnosis and treatment of colon cancer. The expression of tumor-specific antigens is often associated with the progression of neoplastic tissue disorders. Native PRO301 (SEQ ID NO:119) and A33_HUMAN also show a Blast score of 245 and 30% homology at residues 21 to 282 of Fig. 44 with A33_HUMAN, the variation dependent upon how spaces are inserted into the compared sequences. Native PRO301 (SEQ ID NO:119) also has a Blast score of 165 and 29% homology at residues 60 to 255 of Fig. 44 with HS46KDA_1, a human coxsackie and adenovirus receptor protein, also known as cell surface protein HCAR. This region of PRO301 also shows a similar Blast score and homology with HSU90716_1. Expression of such proteins is usually associated with viral infection and therapeutics for the prevention of such infection may be accordingly conceived. As mentioned in the Background, the expression of viral receptors is often associated with neoplastic tumors.

Therapeutic uses for the PRO234 polypeptides of the invention includes treatments associated with leukocyte homing or the interaction between leukocytes and the endothelium during an inflammatory response. Examples

include asthma, rheumatoid arthritis, psoriasis and multiple sclerosis.

Since the PRO231 polypeptide and nucleic acid encoding it possess sequence homology to a putative acid phosphatase and its encoding nucleic acid, probes based upon the PRO231 nucleotide sequence may be employed to identify other novel phosphatase proteins. Soluble forms of the PRO231 polypeptide may be employed as antagonists of membrane bound PRO231 activity both *in vitro* and *in vivo*. PRO231 polypeptides may be employed in screening assays designed to identify agonists or antagonists of the native PRO231 polypeptide, wherein such assays may take the form of any conventional cell-type or biochemical binding assay. Moreover, the PRO231 polypeptide may serve as a molecular marker for the tissues in which the polypeptide is specifically expressed.

PRO229 polypeptides can be fused with peptides of interest to determine whether the fusion peptide has an increased half-life over the peptide of interest. The PRO229 polypeptides can be used accordingly to increase the half-life of polypeptides of interest. Portions of PRO229 which cause the increase in half-life are an embodiment of the invention herein.

PRO238 can be used in assays which measure its ability to reduce substrates, including oxygen and Aceyl-CoA, and particularly, measure PRO238's ability to produce oxygen free radicals. This is done by using assays which have been previously described. PRO238 can further be used to assay for candidates which block, reduce or reverse its reducing abilities. This is done by performing side by side assays where candidates are added in one assay having PRO238 and a substrate to reduce, and not added in another assay, being the same but for the lack of the presence of the candidate.

PRO233 polypeptides and portions thereof which have homology to reductase may also be useful for *in vivo* therapeutic purposes, as well as for various other applications. The identification of novel reductase proteins and related molecules may be relevant to a number of human disorders such as inflammatory disease, organ failure, atherosclerosis, cardiac injury, infertility, birth defects, premature aging, AIDS, cancer, diabetic complications and mutations in general. Given that oxygen free radicals and antioxidants appear to play important roles in a number of disease processes, the identification of new reductase proteins and reductase-like molecules is of special importance in that such proteins may serve as potential therapeutics for a variety of different human disorders. Such polypeptides may also play important roles in biotechnological and medical research, as well as various industrial applications. As a result, there is particular scientific and medical interest in new molecules, such as PRO233.

The PRO223 polypeptides of the present invention which exhibit serine carboxypeptidease activity may be employed *in vivo* for therapeutic purposes as well as for *in vitro* purposes. Those of ordinary skill in the art will well know how to employ PRO223 polypeptides for such uses.

PRO235 polypeptides and portions thereof which may be involved in cell adhesion are also useful for *in vivo* therapeutic purposes, as well as for various *in vitro* applications. In addition, PRO235 polypeptides and portions thereof may have therapeutic applications in disease states which involve cell adhesion. Given the physiological importance of cell adhesion mechanisms *in vivo*, efforts are currently being under taken to identify new, native proteins which are involved in cell adhesion. Therefore, peptides having homology to plexin are of particular interest to the scientific and medical communities.

Because the PRO236 and PRO262 polypeptides disclosed herein are homologous to various known β-galactosidase proteins, the PRO236 and PRO262 polypeptides disclosed herein will find use in conjugates of

As mentioned previously, fibroblast growth factors can act upon cells in both a mitogenic and non-mitogenic manner. These factors are mitogenic for a wide variety of normal diploid mesoderm-derived and neural crest-derived cells, inducing granulosa cells, adrenal cortical cells, chrondrocytes, myoblasts, corneal and vascular endothelial cells (bovine or human), vascular smooth muscle cells, lens, retina and prostatic epithelial cells, oligodendrocytes, astrocytes, chrondocytes, myoblasts and osteoblasts. Antibodies to these factors can be generated to modulate such effects.

Non-mitogenic actions of fibroblast growth factors include promotion of cell migration into a wound area (chemotaxis), initiation of new blood vessel formulation (angiogenesis), modulation of nerve regeneration and survival (neurotrophism), modulation of endocrine functions, and stimulation or suppression of specific cellular protein expression, extracellular matrix production and cell survival. Baird, A. & Bohlen, P., *Handbook of Exp. Phrmacol.* 95(1): 369-418 (1990). These properties provide a basis for using fibroblast growth factors in therapeutic approaches to accelerate wound healing, nerve repair, collateral blood vessel formation, and the like. For example, fibroblast growth factors, have been suggested to minimize myocardium damage in heart disease and surgery (U.S.P. 4,378,437). Antibodies to these factors can be generated to modulate such effects.

Therapeutic indications for PRO214 polypeptides include disorders associated with the preservation and maintenance of gastrointestinal mucosa and the repair of acute and chronic mucosal lesions (e.g., enterocolitis, Zollinger-Ellison syndrome, gastrointestinal ulceration and congenital microvillus atrophy), skin diseases associated with abnormal keratinocyte differentiation (e.g., psoriasis, epithelial cancers such as lung squamous cell carcinoma, epidermoid carcinoma of the vulva and gliomas.

Anti-PRO317 antibodies find use in anti-tumor indications if they are angiostatic, or in coronary ischemic indications if they are angiogenic.

Native PRO301 (SEQ ID NO:119) has a Blast score of 246 and 30% homology at residues 24 to 282 of Fig. 44 with A33_HUMAN, an A33 antigen precursor. A33 antigen precursor, as explained in the Background is a tumor-specific antigen, and as such, is a recognized marker and therapeutic target for the diagnosis and treatment of colon cancer. The expression of tumor-specific antigens is often associated with the progression of neoplastic tissue disorders. Native PRO301 (SEQ ID NO:119) and A33_HUMAN also show a Blast score of 245 and 30% homology at residues 21 to 282 of Fig. 44 with A33_HUMAN, the variation dependent upon how spaces are inserted into the compared sequences. Native PRO301 (SEQ ID NO:119) also has a Blast score of 165 and 29% homology at residues 60 to 255 of Fig. 44 with HS46KDA_1, a human coxsackie and adenovirus receptor protein, also known as cell surface protein HCAR. This region of PRO301 also shows a similar Blast score and homology with HSU90716_1. Expression of such proteins is usually associated with viral infection and therapeutics for the prevention of such infection may be accordingly conceived. Accordingly, antibodies to the above identified antigens and receptors have therapeutic potential as diagnostic and treatment techniques.

Therapeutic uses for the PRO234 polypeptides of the invention includes treatments associated with leukocyte homing or the interaction between leukocytes and the endothelium during an inflammatory response. Examples include asthma, rheumatoid arthritis, psoriasis and multiple sclerosis.

Cancer-associated or specific antigens permit the creation of tumor or cancer specific monoclonal antibodies (mAbs) which are specific to such tumor antigens. Such mAbs, which can distinguish between normal and cancerous

density lipoprotein receptors, apolipoprotein E receptor and chicken oocyte receptors P95. Based on a BLAST and FastA sequence alignment analysis of the full-length sequence, PRO224 has amino acid identity to portions of these proteins in the range from 28% to 45%, and overall identity with these proteins in the range from 33% to 39%.

EXAMPLE 21: Isolation of cDNA Clones Encoding Human PRO222

A consensus DNA sequence was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence is designated herein as DNA28771. Based on the DNA28771 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO222.

A pair of PCR primers (forward and reverse) were synthesized:

forward PCR primer 5'-ATCTCCTATCGCTGCTTTCCCGG-3' (SEQ ID NO:133)

reverse PCR primer 5'-AGCCAGGATCGCAGTAAAACTCC-3' (SEQ ID NO:134)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA28771 sequence which had the following nucleotide sequence:

hybridization probe

5'-ATTTAAACTTGATGGGTCTGCGTATCTTGAGTGCTTACAAAACCTTATCT-3' (SEQ ID NO:135)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pair identified above. A positive library was then used to isolate clones encoding the PRO222 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal kidney tissue.

DNA sequencing of the clones isolated as described above gave the full-length DNA sequence for PRO222 [herein designated as UNQ196 (DNA33107-1135)] and the derived protein sequence for PRO222.

The entire nucleotide sequence of UNQ196 (DNA33107-1135) is shown in Figure 47 (SEQ ID NO:131). Clone UNQ196 (DNA33107-1135) contains a single open reading frame with an apparent translational initiation site at nucleotide positions 159-161 and ending at the stop codon at nucleotide positions 1629-1631 (Fig. 47; SEQ ID NO:131). The predicted polypeptide precursor is 490 amino acids long (Fig. 48). Clone UNQ196 (DNA33107-1135) has been deposited with ATCC and is assigned ATCC deposit no. ATCC 209251.

Based on a BLAST and FastA sequence alignment analysis of the full-length sequence, PRO222 shows amino acid sequence identity to mouse complement factor h precursor (25-26%), complement receptor (27-29%), mouse complement C3b receptor type 2 long form precursor (25-47%) and human hypothetical protein kiaa0247 (40%).

EXAMPLE 22: Isolation of cDNA clones Encoding PRO234

A consensus DNA sequence was assembled (DNA30926) using phrap as described in Example 1 above. Based on this consensus sequence, oligonucleotides were synthesized: 1) to identify by PCR a cDNA library that contained the sequence of interest, and 2) for use as probes to isolate a clone of the full-length coding sequence.

RNA for the construction of the cDNA libraries was isolated using standard isolation protocols, e.g., Ausubel *et al.*, *Current Protocols in Molecular Biology*, from tissue or cell line sources or it was purchased from

commercial sources (e.g., Clontech). The cDNA libraries used to isolate the cDNA clones were constructed by standard methods (e.g., Ausubel *et al.*) using commercially available reagents (e.g., Invitrogen). This library was derived from 22 week old fetal brain tissue.

A cDNA clone was sequenced in its entirety. The entire nucleotide sequence of PRO234 is shown in Figure 49 (SEQ ID NO:136). The predicted polypeptide precursor is 382 amino acids long and has a calculated molecular weight of approximately 43.1 kDa.

The oligonucleotide sequences used in the above procedure were the following:

30926.p (OLI826) (SEQ ID NO:138): 5'-GTTCATTGAAAACCTCTTGCCATCT GATGGTGACTTCTGGATTGGGCTCA-3'

30926.f (OLI827) (SEQ ID NO:139): 5'-AAGCCAAAGAAGCCTGCAGGAGGG-3'

30926.r (OLI828) (SEQ ID NO:140): 5'-CAGTCCAAGCATAAAGGTCCTGGC-3'

EXAMPLE 23: Isolation of cDNA Clones Encoding Human PRO231

A consensus DNA sequence was assembled relative to the other identified EST sequences as described in Example 1 above, wherein the consensus sequence was designated herein as DNA30933. Based on the DNA30933 consensus sequence, oligonucleotides were synthesized to identify by PCR a cDNA library that contained the sequence of interest and for use as probes to isolate a clone of the full-length coding sequence for PRO231.

Three PCR primers (two forward and one reverse) were synthesized:

forward PCR primer 1 5'-CCAACTACCAAAGCTGCTGGAGCC-3' (SEQ ID NO:143)

forward PCR primer 2 5'-GCAGCTCTATTACCACGGGAAGGA-3' (SEQ ID NO:144)

reverse PCR primer 5'-TCCTTCCCGTGGTAATAGAGCTGC-3' (SEQ ID NO:145)

Additionally, a synthetic oligonucleotide hybridization probe was constructed from the consensus DNA30933 sequence which had the following nucleotide sequence

hybridization probe

5'-GGCAGAGAACCAGAGGCCGGAGGAGACTGCCTCTTTACAGCCAGG-3' (SEQ ID NO:146)

In order to screen several libraries for a source of a full-length clone, DNA from the libraries was screened by PCR amplification with the PCR primer pairs identified above. A positive library was then used to isolate clones encoding the PRO231 gene using the probe oligonucleotide and one of the PCR primers.

RNA for construction of the cDNA libraries was isolated from human fetal liver tissue. DNAsequencing of the clones isolated as described above gave the full-length DNA sequence for PRO231 [herein designated as UNQ205 (DNA34434-1139)] and the derived protein sequence for PRO231.

The entire nucleotide sequence of UNQ205 (DNA34434-1139) is shown in Figure 51 (SEQ ID NO:141). Clone UNQ205 (DNA34434-1139) contains a single open reading frame with an apparent translational initiation site at nucleotide positions 173-175 and ending at the stop codon at nucleotide positions 1457-1459 (Fig. 51; SEQ ID NO:141). The predicted polypeptide precursor is 428 amino acids long (Fig. 52). Clone UNQ205 (DNA34434-1139) has been deposited with ATCC on September 16, 1997 and is assigned ATCC deposit no. ATCC 209252.

Analysis of the amino acid sequence of the full-length PRO231 suggests that it possesses 30% and 31% amino acid identity with the human and rat prostatic acid phosphatase precursor proteins, respectively.

most compact with their hydrophobic interiors shielded from interaction with the reversed phase resin. Aggregated species are usually eluted at higher acetonitrile concentrations. In addition to resolving misfolded forms of proteins from the desired form, the reversed phase step also removes endotoxin from the samples.

Fractions containing the desired folded PRO187, PRO317, PRO301, PRO224 and PRO238 proteins, respectively, were pooled and the acetonitrile removed using a gentle stream of nitrogen directed at the solution. Proteins were formulated into 20 mM Hepes, pH 6.8 with 0.14 M sodium chloride and 4% mannitol by dialysis or by gel filtration using G25 Superfine (Pharmacia) resins equilibrated in the formulation buffer and sterile filtered.

EXAMPLE 54: Expression of PRO Polypeptides in Mammalian Cells

This example illustrates preparation of a glycosylated form of a desired PRO polypeptide by recombinant expression in mammalian cells.

The vector, pRK5 (see EP 307,247, published March 15, 1989), is employed as the expression vector. Optionally, the PRO polypeptide-encoding DNA is ligated into pRK5 with selected restriction enzymes to allow insertion of the PRO polypeptide DNA using ligation methods such as described in Sambrook et al., supra. The resulting vector is called pRK5-PRO polypeptide.

In one embodiment, the selected host cells may be 293 cells. Human 293 cells (ATCC CCL 1573) are grown to confluence in tissue culture plates in medium such as DMEM supplemented with fetal calf serum and optionally, nutrient components and/or antibiotics. About 10 μg pRK5-PRO polypeptide DNA is mixed with about 1 μg DNA encoding the VA RNA gene [Thimmappaya et al., Cell, 31:543 (1982)] and dissolved in 500 μl of 1 mM Tris-HCl, 0.1 mM EDTA, 0.227 M $CaCl_2$. To this mixture is added, dropwise, 500 μl of 50 mM HEPES (pH 7.35), 280 mM NaCl, 1.5 mM $NaPO_4$, and a precipitate is allowed to form for 10 minutes at 25°C. The precipitate is suspended and added to the 293 cells and allowed to settle for about four hours at 37°C. The culture medium is aspirated off and 2 ml of 20% glycerol in PBS is added for 30 seconds. The 293 cells are then washed with serum free medium, fresh medium is added and the cells are incubated for about 5 days.

Approximately 24 hours after the transfections, the culture medium is removed and replaced with culture medium (alone) or culture medium containing 200 μCi/ml ^{35}S-cysteine and 200 μCi/ml ^{35}S-methionine. After a 12 hour incubation, the conditioned medium is collected, concentrated on a spin filter, and loaded onto a 15% SDS gel. The processed gel may be dried and exposed to film for a selected period of time to reveal the presence of PRO polypeptide. The cultures containing transfected cells may undergo further incubation (in serum free medium) and the medium is tested in selected bioassays.

In an alternative technique, PRO polypeptide may be introduced into 293 cells transiently using the dextran sulfate method described by Somparyrac et al., Proc. Natl. Acad. Sci., 12:7575 (1981). 293 cells are grown to maximal density in a spinner flask and 700 μg pRK5-PRO polypeptide DNA is added. The cells are first concentrated from the spinner flask by centrifugation and washed with PBS. The DNA-dextran precipitate is incubated on the cell pellet for four hours. The cells are treated with 20% glycerol for 90 seconds, washed with tissue culture medium, and re-introduced into the spinner flask containing tissue culture medium, 5 μg/ml bovine insulin and 0.1 μg/ml bovine transferrin. After about four days, the conditioned media is centrifuged and filtered to remove cells and debris. The sample containing expressed PRO polypeptide can then be concentrated and purified by any selected

method, such as dialysis and/or column chromatography.

In another embodiment, PRO polypeptides can be expressed in CHO cells. The pRK5-PRO polypeptide can be transfected into CHO cells using known reagents such as $CaPO_4$ or DEAE-dextran. As described above, the cell cultures can be incubated, and the medium replaced with culture medium (alone) or medium containing a radiolabel such as ^{35}S-methionine. After determining the presence of PRO polypeptide, the culture medium may be replaced with serum free medium. Preferably, the cultures are incubated for about 6 days, and then the conditioned medium is harvested. The medium containing the expressed PRO polypeptide can then be concentrated and purified by any selected method.

Epitope-tagged PRO polypeptide may also be expressed in host CHO cells. The PRO polypeptide may be subcloned out of the pRK5 vector. The subclone insert can undergo PCR to fuse in frame with a selected epitope tag such as a poly-his tag into a Baculovirus expression vector. The poly-his tagged PRO polypeptide insert can then be subcloned into a SV40 driven vector containing a selection marker such as DHFR for selection of stable clones. Finally, the CHO cells can be transfected (as described above) with the SV40 driven vector. Labeling may be performed, as described above, to verify expression. The culture medium containing the expressed poly-His tagged PRO polypeptide can then be concentrated and purified by any selected method, such as by Ni^{2+}-chelate affinity chromatography.

PRO211, PRO217, PRO230, PRO219, PRO245, PRO221, PRO258, PRO301, PRO224, PRO222, PRO234, PRO229, PRO223, PRO328 and PRO332 were successfully expressed in CHO cells by both a transient and a stable expression procedure. In addition, PRO232, PRO265, PRO246, PRO228, PRO227, PRO220, PRO266, PRO269, PRO287, PRO214, PRO231, PRO233, PRO238, PRO244, PRO235, PRO236, PRO262, PRO239, PRO257, PRO260, PRO263, PRO270, PRO271, PRO272, PRO294, PRO295, PRO293, PRO247, PRO303 and PRO268 were successfully transiently expressed in CHO cells.

Stable expression in CHO cells was performed using the following procedure. The proteins were expressed as an IgG construct (immunoadhesin), in which the coding sequences for the soluble forms (e.g. extracellular domains) of the respective proteins were fused to an IgG1 constant region sequence containing the hinge, CH2 and CH2 domains and/or is a poly-His tagged form.

Following PCR amplification, the respective DNAs were subcloned in a CHO expression vector using standard techniques as described in Ausubel et al., *Current Protocols of Molecular Biology*, Unit 3.16, John Wiley and Sons (1997). CHO expression vectors are constructed to have compatible restriction sites 5' and 3' of the DNA of interest to allow the convenient shuttling of cDNA's. The vector used expression in CHO cells is as described in Lucas *et al.*, *Nucl. Acids Res.* 24: 9 (1774-1779 (1996), and uses the SV40 early promoter/enhancer to drive expression of the cDNA of interest and dihydrofolate reductase (DHFR). DHFR expression permits selection for stable maintenance of the plasmid following transfection.

Twelve micrograms of the desired plasmid DNA were introduced into approximately 10 million CHO cells using commercially available transfection reagents Superfect® (Quiagen), Dosper® or Fugene® (Boehringer Mannheim). The cells were grown and described in Lucas *et al.*, supra. Approximately 3 x 10^{-7} cells are frozen in an ampule for further growth and production as described below.

The ampules containing the plasmid DNA were thawed by placement into water bath and mixed by vortexing. The contents were pipetted into a centrifuge tube containing 10 mLs of media and centrifuged at 1000 rpm for 5 minutes. The supernatant was aspirated and the cells were resuspended in 10 mL of selective media (0.2 μm filtered PS20 with 5% 0.2 μm diafiltered fetal bovine serum). The cells were then aliquoted into a 100 mL spinner containing 90 mL of selective media. After 1-2 days, the cells were transferred into a 250 mL spinner filled with 150 mL selective growth medium and incubated at 37°C. After another 2-3 days, a 250 mL, 500 mL and 2000 mL spinners were seeded with 3 x 10^5 cells/mL. The cell media was exchanged with fresh media by centrifugation and resuspension in production medium. Although any suitable CHO media may be employed, a production medium described in US Patent No. 5,122,469, issued June 16, 1992 was actually used. 3L production spinner is seeded at 1.2 x 10^6 cells/mL. On day 0, the cell number pH were determined. On day 1, the spinner was sampled and sparging with filtered air was commenced. On day 2, the spinner was sampled, the temperature shifted to 33°C, and 30 mL of 500 g/L glucose and 0.6 mL of 10% antifoam (e.g., 35% polydimethylsiloxane emulsion, Dow Corning 365 Medical Grade Emulsion). Throughout the production, pH was adjusted as necessary to keep at around 7.2. After 10 days, or until viability dropped below 70%, the cell culture was harvested by centrifugtion and filtering through a 0.22 μm filter. The filtrate was either stored at 4°C or immediately loaded onto columns for purification.

For the poly-His tagged constructs, the proteins were purified using a Ni-NTA column (Qiagen). Before purification, imidazole was added to the conditioned media to a concentration of 5 mM. The conditioned media was pumped onto a 6 ml Ni-NTA column equilibrated in 20 mM Hepes, pH 7.4, buffer containing 0.3 M NaCl and 5 mM imidazole at a flow rate of 4-5 ml/min. at 4°C. After loading, the column was washed with additional equilibration buffer and the protein eluted with equilibration buffer containing 0.25 M imidazole. The highly purified protein was subsequently desalted into a storage buffer containing 10 mM Hepes, 0.14 M NaCl and 4% mannitol, pH 6.8, with a 25 ml G25 Superfine (Pharmacia) column and stored at -80°C.

Immunoadhesin (Fc containing) constructs of were purified from the conditioned media as follows. The conditioned medium was pumped onto a 5 ml Protein A column (Pharmacia) which had been equilibrated in 20 mM Na phosphate buffer, pH 6.8. After loading, the column was washed extensively with equilibration buffer before elution with 100 mM citric acid, pH 3.5. The eluted protein was immediately neutralized by collecting 1 ml fractions into tubes containing 275 μL of 1 M Tris buffer, pH 9. The highly purified protein was subsequently desalted into storage buffer as described above for the poly-His tagged proteins. The homogeneity was assessed by SDS polyacrylamide gels and by N-terminal amino acid sequencing by Edman degradation.

PRO211, PRO217, PRO230, PRO232, PRO187, PRO265, PRO219, PRO246, PRO228, PRO533, PRO245, PRO221, PRO227, PRO220, PRO258, PRO266, PRO269, PRO287, PRO214, PRO317, PRO301, PRO224, PRO222, PRO234, PRO231, PRO229, PRO233, PRO238, PRO223, PRO235, PRO236, PRO262, PRO239, PRO257, PRO260, PRO263, PRO270, PRO271, PRO272, PRO294, PRO295, PRO293, PRO247, PRO304, PRO302, PRO307, PRO303, PRO343, PRO328, PRO326, PRO331, PRO332, PRO334, PRO346, PRO268, PRO330, PRO310 and PRO339 were also successfully transiently expressed in COS cells.

EXAMPLE 55: Expression of PRO Polypeptides in Yeast

The following method describes recombinant expression of a desired PRO polypeptide in yeast.

First, yeast expression vectors are constructed for intracellular production or secretion of PRO polypeptides from the ADH2/GAPDH promoter. DNA encoding a desired PRO polypeptide, a selected signal peptide and the promoter is inserted into suitable restriction enzyme sites in the selected plasmid to direct intracellular expression of the PRO polypeptide. For secretion, DNA encoding the PRO polypeptide can be cloned into the selected plasmid, together with DNA encoding the ADH2/GAPDH promoter, the yeast alpha-factor secretory signal/leader sequence, and linker sequences (if needed) for expression of the PRO polypeptide.

Yeast cells, such as yeast strain AB110, can then be transformed with the expression plasmids described above and cultured in selected fermentation media. The transformed yeast supernatants can be analyzed by precipitation with 10% trichloroacetic acid and separation by SDS-PAGE, followed by staining of the gels with Coomassie Blue stain.

Recombinant PRO polypeptide can subsequently be isolated and purified by removing the yeast cells from the fermentation medium by centrifugation and then concentrating the medium using selected cartridge filters. The concentrate containing the PRO polypeptide may further be purified using selected column chromatography resins.

EXAMPLE 56: Expression of PRO Polypeptides in Baculovirus-Infected Insect Cells

The following method describes recombinant expression of PRO polypeptides in Baculovirus-infected insect cells.

The desired PRO polypeptide is fused upstream of an epitope tag contained with a baculovirus expression vector. Such epitope tags include poly-his tags and immunoglobulin tags (like Fc regions of IgG). A variety of plasmids may be employed, including plasmids derived from commercially available plasmids such as pVL1393 (Novagen). Briefly, the PRO polypeptide or the desired portion of the PRO polypeptide (such as the sequence encoding the extracellular domain of a transmembrane protein) is amplified by PCR with primers complementary to the 5' and 3' regions. The 5' primer may incorporate flanking (selected) restriction enzyme sites. The product is then digested with those selected restriction enzymes and subcloned into the expression vector.

Recombinant baculovirus is generated by co-transfecting the above plasmid and BaculoGold™ virus DNA (Pharmingen) into *Spodoptera frugiperda* ("Sf9") cells (ATCC CRL 1711) using lipofectin (commercially available from GIBCO-BRL). After 4-5 days of incubation at 28°C, the released viruses are harvested and used for further amplifications. Viral infection and protein expression is performed as described by O'Reilley et al., Baculovirus expression vectors: A laboratory Manual, Oxford: Oxford University Press (1994).

Expressed poly-his tagged PRO polypeptide can then be purified, for example, by Ni^{2+}-chelate affinity chromatography as follows. Extracts are prepared from recombinant virus-infected Sf9 cells as described by Rupert *et al., Nature*, 362:175-179 (1993). Briefly, Sf9 cells are washed, resuspended in sonication buffer (25 mL Hepes, pH 7.9; 12.5 mM $MgCl_2$; 0.1 mM EDTA; 10% Glycerol; 0.1% NP-40; 0.4 M KCl), and sonicated twice for 20 seconds on ice. The sonicates are cleared by centrifugation, and the supernatant is diluted 50-fold in loading buffer (50 mM phosphate, 300 mM NaCl, 10% Glycerol, pH 7.8) and filtered through a 0.45 μm filter. A Ni^{2+}-NTA agarose column (commercially available from Qiagen) is prepared with a bed volume of 5 mL, washed with 25 mL

of water and equilibrated with 25 mL of loading buffer. The filtered cell extract is loaded onto the column at 0.5 mL per minute. The column is washed to baseline A_{280} with loading buffer, at which point fraction collection is started. Next, the column is washed with a secondary wash buffer (50 mM phosphate; 300 mM NaCl, 10% Glycerol, pH 6.0), which elutes nonspecifically bound protein. After reaching A_{280} baseline again, the column is developed with a 0 to 500 mM Imidazole gradient in the secondary wash buffer. One mL fractions are collected and analyzed by SDS-PAGE and silver staining or western blot with Ni^{2+}-NTA-conjugated to alkaline phosphatase (Qiagen). Fractions containing the eluted His_{10}-tagged PRO polypeptide are pooled and dialyzed against loading buffer.

Alternatively, purification of the IgG tagged (or Fc tagged) PRO polypeptide can be performed using known chromatography techniques, including for instance, Protein A or protein G column chromatography.

PRO211, PRO217, PRO230, PRO187, PRO265, PRO246, PRO228, PRO533, PRO245, PRO221, PRO220, PRO258, PRO266, PRO269, PRO287, PRO214, PRO301, PRO224, PRO222, PRO234, PRO231, PRO229, PRO235, PRO239, PRO257, PRO272, PRO294, PRO295, PRO328, PRO326, PRO331, PRO334, PRO346 and PRO310 were successfully expressed in baculovirus infected Sf9 or high5 insect cells. While the expression was actually performed in a 0.5-2 L scale, it can be readily scaled up for larger (e.g. 8 L) preparations. The proteins were expressed as an IgG construct (immunoadhesin), in which the protein extracellular region was fused to an IgG1 constant region sequence containing the hinge, CH2 and CH3 domains and/or in poly-His tagged forms.

Following PCR amplification, the respective coding sequences were subcloned into a baculovirus expression vector (pb.PH.IgG for IgG fusions and pb.PH.His.c for poly-His tagged proteins), and the vector and Baculogold® baculovirus DNA (Pharmingen) were co-transfected into 105 *Spodoptera frugiperda* ("Sf9") cells (ATCC CRL 1711), using Lipofectin (Gibco BRL). pb.PH.IgG and pb.PH.His are modifications of the commercially available baculovirus expression vector pVL1393 (Pharmingen), with modified polylinker regions to include the His or Fc tag sequences. The cells were grown in Hink's TNM-FH medium supplemented with 10% FBS (Hyclone). Cells were incubated for 5 days at 28°C. The supernatant was harvested and subsequently used for the first viral amplification by infecting Sf9 cells in Hink's TNM-FH medium supplemented with 10% FBS at an approximate multiplicity of infection (MOI) of 10. Cells were incubated for 3 days at 28°C. The supernatant was harvested and the expression of the constructs in the baculovirus expression vector was determined by batch binding of 1 ml of supernatant to 25 mL of Ni-NTA beads (QIAGEN) for histidine tagged proteins or Protein-A Sepharose CL-4B beads (Pharmacia) for IgG tagged proteins followed by SDS-PAGE analysis comparing to a known concentration of protein standard by Coomassie blue staining.

The first viral amplification supernatant was used to infect a spinner culture (500 ml) of Sf9 cells grown in ESF-921 medium (Expression Systems LLC) at an approximate MOI of 0.1. Cells were incubated for 3 days at 28°C. The supernatant was harvested and filtered. Batch binding and SDS-PAGE analysis was repeated, as necessary, until expression of the spinner culture was confirmed.

The conditioned medium from the transfected cells (0.5 to 3 L) was harvested by centrifugation to remove the cells and filtered through 0.22 micron filters. For the poly-His tagged constructs, the protein construct were purified using a Ni-NTA column (Qiagen). Before purification, imidazole was added to the conditioned media to a concentration of 5 mM. The conditioned media were pumped onto a 6 ml Ni-NTA column equilibrated in 20 mM Hepes, pH 7.4, buffer containing 0.3 M NaCl and 5 mM imidazole at a flow rate of 4-5 ml/min. at 4°C. After

loading, the column was washed with additional equilibration buffer and the protein eluted with equilibration buffer containing 0.25 M imidazole. The highly purified protein was subsequently desalted into a storage buffer containing 10 mM Hepes, 0.14 M NaCl and 4% mannitol, pH 6.8, with a 25 ml G25 Superfine (Pharmacia) column and stored at -80°C.

Immunoadhesin (Fc containing) constructs of proteins were purified from the conditioned media as follows. The conditioned media were pumped onto a 5 ml Protein A column (Pharmacia) which had been equilibrated in 20 mM Na phosphate buffer, pH 6.8. After loading, the column was washed extensively with equilibration buffer before elution with 100 mM citric acid, pH 3.5. The eluted protein was immediately neutralized by collecting 1 ml fractions into tubes containing 275 mL of 1 M Tris buffer, pH 9. The highly purified protein was subsequently desalted into storage buffer as described above for the poly-His tagged proteins. The homogeneity of the proteins was verified by SDS polyacrylamide gel (PEG) electrophoresis and N-terminal amino acid sequencing by Edman degradation.

EXAMPLE 57: Preparation of Antibodies that Bind to PRO Polypeptides

This example illustrates preparation of monoclonal antibodies which can specifically bind to a PRO polypeptide.

Techniques for producing the monoclonal antibodies are known in the art and are described, for instance, in Goding, supra. Immunogens that may be employed include purified PRO polypeptide, fusion proteins containing the PRO polypeptide, and cells expressing recombinant PRO polypeptide on the cell surface. Selection of the immunogen can be made by the skilled artisan without undue experimentation.

Mice, such as Balb/c, are immunized with the PRO polypeptide immunogen emulsified in complete Freund's adjuvant and injected subcutaneously or intraperitoneally in an amount from 1-100 micrograms. Alternatively, the immunogen is emulsified in MPL-TDM adjuvant (Ribi Immunochemical Research, Hamilton, MT) and injected into the animal's hind foot pads. The immunized mice are then boosted 10 to 12 days later with additional immunogen emulsified in the selected adjuvant. Thereafter, for several weeks, the mice may also be boosted with additional immunization injections. Serum samples may be periodically obtained from the mice by retro-orbital bleeding for testing in ELISA assays to detect anti-PRO polypeptide antibodies.

After a suitable antibody titer has been detected, the animals "positive" for antibodies can be injected with a final intravenous injection of PRO polypeptide. Three to four days later, the mice are sacrificed and the spleen cells are harvested. The spleen cells are then fused (using 35% polyethylene glycol) to a selected murine myeloma cell line such as P3X63AgU.1, available from ATCC, No. CRL 1597. The fusions generate hybridoma cells which can then be plated in 96 well tissue culture plates containing HAT (hypoxanthine, aminopterin, and thymidine) medium to inhibit proliferation of non-fused cells, myeloma hybrids, and spleen cell hybrids.

The hybridoma cells will be screened in an ELISA for reactivity against the PRO polypeptide. Determination of "positive" hybridoma cells secreting the desired monoclonal antibodies against the PRO polypeptide is within the skill in the art.

The positive hybridoma cells can be injected intraperitoneally into syngeneic Balb/c mice to produce ascites containing the anti-PRO polypeptide monoclonal antibodies. Alternatively, the hybridoma cells can be grown in tissue culture flasks or roller bottles. Purification of the monoclonal antibodies produced in the ascites can be accomplished

analyze the tissue distribution of transcription, identify and localize viral infection, follow changes in specific mRNA synthesis and aid in chromosome mapping.

In situ hybridization was performed following an optimized version of the protocol by Lu and Gillett, Cell Vision 1:169-176 (1994), using PCR-generated ^{33}P-labeled riboprobes. Briefly, formalin-fixed, paraffin-embedded human tissues were sectioned, deparaffinized, deproteinated in proteinase K (20 g/ml) for 15 minutes at 37°C, and further processed for *in situ* hybridization as described by Lu and Gillett, *supra*. A [^{33}-P] UTP-labeled antisense riboprobe was generated from a PCR product and hybridized at 55°C overnight. The slides were dipped in Kodak NTB2 nuclear track emulsion and exposed for 4 weeks.

^{33}P-Riboprobe synthesis

6.0 μl (125 mCi) of ^{33}P-UTP (Amersham BF 1002, SA < 2000 Ci/mmol) were speed vac dried. To each tube containing dried ^{33}P-UTP, the following ingredients were added:

2.0 μl 5x transcription buffer
1.0 μl DTT (100 mM)
2.0 μl NTP mix (2.5 mM : 10 μ; each of 10 mM GTP, CTP & ATP + 10 μl H_2O)
1.0 μl UTP (50 μM)
1.0 μl Rnasin
1.0 μl DNA template (1μg)
1.0 μl H_2O
1.0 μl RNA polymerase (for PCR products T3 = AS, T7 = S, usually)

The tubes were incubated at 37°C for one hour. 1.0 μl RQ1 DNase were added, followed by incubation at 37°C for 15 minutes. 90 μl TE (10 mM Tris pH 7.6/1mM EDTA pH 8.0) were added, and the mixture was pipetted onto DE81 paper. The remaining solution was loaded in a Microcon-50 ultrafiltration unit, and spun using program 10 (6 minutes). The filtration unit was inverted over a second tube and spun using program 2 (3 minutes). After the final recovery spin, 100 μl TE were added. 1 μl of the final product was pipetted on DE81 paper and counted in 6 ml of Biofluor II.

The probe was run on a TBE/urea gel. 1-3 μl of the probe or 5 μl of RNA Mrk III were added to 3 μl of loading buffer. After heating on a 95°C heat block for three minutes, the gel was immediately placed on ice. The wells of gel were flushed, the sample loaded, and run at 180-250 volts for 45 minutes. The gel was wrapped in saran wrap and exposed to XAR film with an intensifying screen in -70°C freezer one hour to overnight.

^{33}P-Hybridization

A. Pretreatment of frozen sections

The slides were removed from the freezer, placed on aluminium trays and thawed at room temperature for 5 minutes. The trays were placed in 55°C incubator for five minutes to reduce condensation. The slides were fixed for 10 minutes in 4% paraformaldehyde on ice in the fume hood, and washed in 0.5 x SSC for 5 minutes, at room temperature (25 ml 20 x SSC + 975 ml SQ H_2O). After deproteination in 0.5 μg/ml proteinase K for 10 minutes at 37°C (12.5 μl of 10 mg/ml stock in 250 ml prewarmed RNase-free RNAse buffer), the sections were washed in 0.5 x SSC for 10 minutes at room temperature. The sections were dehydrated in 70%, 95%, 100% ethanol, 2 minutes each.

B. Pretreatment of paraffin-embedded sections

The slides were deparaffinized, placed in SQ H_2O, and rinsed twice in 2 x SSC at room temperature, for 5 minutes each time. The sections were deproteinated in 20 μg/ml proteinase K (500 μl of 10 mg/ml in 250 ml RNase-free RNase buffer; 37°C, 15 minutes) - human embryo, or 8 x proteinase K (100 μl in 250 ml Rnase buffer, 37°C, 30 minutes) - formalin tissues. Subsequent rinsing in 0.5 x SSC and dehydration were performed as described above.

C. Prehybridization

The slides were laid out in a plastic box lined with Box buffer (4 x SSC, 50% formamide) - saturated filter paper. The tissue was covered with 50 μl of hybridization buffer (3.75g Dextran Sulfate + 6 ml SQ H_2O), vortexed and heated in the microwave for 2 minutes with the cap loosened. After cooling on ice, 18.75 ml formamide, 3.75 ml 20 x SSC and 9 ml SQ H_2O were added, the tissue was vortexed well, and incubated at 42°C for 1-4 hours.

D. Hybridization

1.0 x 10^6 cpm probe and 1.0 μl tRNA (50 mg/ml stock) per slide were heated at 95°C for 3 minutes. The slides were cooled on ice, and 48 μl hybridization buffer were added per slide. After vortexing, 50 μl ^{33}P mix were added to 50 μl prehybridization on slide. The slides were incubated overnight at 55°C.

E. Washes

Washing was done 2 x 10 minutes with 2xSSC, EDTA at room temperature (400 ml 20 x SSC + 16 ml 0.25M EDTA, V_f=4L), followed by RNaseA treatment at 37°C for 30 minutes (500 μl of 10 mg/ml in 250 ml Rnase buffer = 20 μg/ml), The slides were washed 2 x 10 minutes with 2 x SSC, EDTA at room temperature. The stringency wash conditions were as follows: 2 hours at 55°C, 0.1 x SSC, EDTA (20 ml 20 x SSC + 16 ml EDTA, V_f=4L).

F. Oligonucleotides

In situ analysis was performed on a variety of DNA sequences disclosed herein. The oligonucleotides employed for these analyses are as follows.

(1) DNA33094-1131 (PRO217)

p1 5'-GGATTCTAATACGACTCACTATAGGGCTCAGAAAAGCGCAACAGAGAA-3' (SEQ ID NO:348)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGATGTCTTCCATGCCAACCTTC-3' (SEQ ID NO:349)

(2) DNA33223-1136 (PRO230)

p1 5'-GGATTCTAATACGACTCACTATAGGGCGGCGATGTCCACTGGGGCTAC-3' (SEQ ID NO:350)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGACGAGGAAGATGGGCGGATGGT-3' (SEQ ID NO:351)

(3) DNA34435-1140 (PRO232)

p1 5'-GGATTCTAATACGACTCACTATAGGGCACCCACGCGTCCGGCTGCTT-3' (SEQ ID NO:352)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGACGGGGGACACCACGGACCAGA-3' (SEQ ID NO:353)

(4) DNA35639-1172 (PRO246)

p1 5'-GGATTCTAATACGACTCACTATAGGGCTTGCTGCGGTTTTTGTTCCTG-3' (SEQ ID NO:354)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAGCTGCCGATCCCACTGGTATT-3' (SEQ ID NO:355)

(5) DNA49435-1219 (PRO533)

p1 5'-GGATTCTAATACGACTCACTATAGGGCGGATCCTGGCCGGCCTCTG-3' (SEQ ID NO:356)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAGCCCGGGCATGGTCTCAGTTA-3' (SEQ ID NO:357)

(6) DNA35638-1141 (PRO245)

p1 5'-GGATTCTAATACGACTCACTATAGGGCGGGAAGATGGCGAGGAGGAG-3' (SEQ ID NO:358)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGACCAAGGCCACAAACGGAAATC-3' (SEQ ID NO:359)

(7) DNA33089-1132 (PRO221)

p1 5'-GGATTCTAATACGACTCACTATAGGGCTGTGCTTTCATTCTGCCAGTA-3' (SEQ ID NO:360)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAGGGTACAATTAAGGGGTGGAT-3' (SEQ ID NO:361)

(8) DNA35918-1174 (PRO258)

p1 5'-GGATTCTAATACGACTCACTATAGGGCCCGCCTCGCTCCTGCTCCTG-3' (SEQ ID NO:362)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAGGATTGCCGCGACCCTCACAG-3' (SEQ ID NO:363)

(9) DNA32286-1191 (PRO214)

p1 5'-GGATTCTAATACGACTCACTATAGGGCCCCTCCTGCCTTCCCTGTCC-3' (SEQ ID NO:364)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAGTGGTGGCCGCGATTATCTGC-3' (SEQ ID NO:365)

(10) DNA33221-1133 (PRO224)

p1 5'-GGATTCTAATACGACTCACTATAGGGCGCAGCGATGGCAGCGATGAGG-3' (SEQ ID NO:366)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGACAGACGGGGCAGAGGGAGTG-3' (SEQ ID NO:367)

(11) DNA35557-1137 (PRO234)

p1 5'-GGATTCTAATACGACTCACTATAGGGCCAGGAGGCGTGAGGAGAAAC-3' (SEQ ID NO:368)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGAAAGACATGTCATCGGGAGTGG-3' (SEQ ID NO:369)

(12) DNA33100-1159 (PRO229)

p1 5'-GGATTCTAATACGACTCACTATAGGGCCGGGTGGAGGTGGAACAGAAA-3' (SEQ ID NO:370)

p2 5'-CTATGAAATTAACCCTCACTAAAGGGACACAGACAGAGCCCCATACGC-3' (SEQ ID NO:371)

spinal ganglia. All other tissues negative.

Human fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

Adult human tissues examined: Kidney (normal and end-stage), adrenal, myocardium, aorta, spleen, lymph node, pancreas, lung, skin, eye (inc. retina), bladder, liver (normal, cirrhotic, acute failure).

Non-human primate tissues examined:

Chimp Tissues: Salivary gland, stomach, thyroid, parathyroid, skin, thymus, ovary, lymph node.

Rhesus Monkey Tissues: Cerebral cortex, hippocampus, cerebellum, penis.

(11) DNA35557-1137 (PRO234)

Specific expression over developing motor neurones in ventral aspect of the fetal spinal cord (will develop into ventral horns of spinal cord). All other tissues negative. Possible role in growth, differentiation and/or development of spinal motor neurons.

Fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

Adult tissues examined: Liver, kidney, adrenal, myocardium, aorta, spleen, lymph node, pancreas, lung, skin, cerebral cortex (rm), hippocampus(rm), cerebellum(rm), penis, eye, bladder, stomach, gastric carcinoma, colon, colonic carcinoma and chondrosarcoma. Acetominophen induced liver injury and hepatic cirrhosis

(12) DNA33100-1159 (PRO229)

Striking expression in mononuclear phagocytes (macrophages) of fetal and adult spleen, liver, lymph node and adult thymus (in tingible body macrophages). The highest expression is in the spleen. All other tissues negative. Localisation and homology are entirely consistent with a role as a scavenger receptor for cells of the reticuloendothelial system. Expression also observed in placental mononuclear cells.

Human fetal tissues examined (E12-E16 weeks) include: Placenta, umbilical cord, liver, kidney, adrenals, thyroid, lungs, heart, great vessels, oesophagus, stomach, small intestine, spleen, thymus, pancreas, brain, eye, spinal cord, body wall, pelvis and lower limb.

Adult human tissues examined: Kidney (normal and end-stage), adrenal, myocardium, aorta, spleen, lymph node, gall bladder, pancreas, lung, skin, eye (inc. retina), prostate, bladder, liver (normal, cirrhotic, acute failure).

Non-human primate tissues examined:

Chimp Tissues: Salivary gland, stomach, thyroid, parathyroid, skin, thymus, ovary, lymph node.

Rhesus Monkey Tissues: Cerebral cortex, hippocampus, cerebellum, penis.

(13) DNA34431-1177 (PRO263)

Widepread expression in human fetal tissues and placenta over mononuclear cells, probably macrophages +/- lymphocytes. The cellular distribution follows a perivascular pattern in many tissues. Strong expression also seen

FIGURE 49

CCCACGCGTCCGCTCCGCGCCCTCCCCCCCGCCTCCCGTGCGGTCCGTCGGTGGCCTAGAGA
TGCTGCTGCCGCGGTTGCAGTTGTCGCGCACGCCTCTGCCCGCCAGCCCGCTCCACCGCCGT
AGCGCCCGAGTGTCGGGGGGCGCACCCGAGTCGGGCCATGAGGCCGGGAACCGCGCTACAGG
CCGTGCTGCTGGCCGTGCTGCTGGTGGGGCTGCGGGCCGCGACGGGTCGCCTGCTGAGTGCC
TCGGATTTGGACCTCAGAGGAGGGCAGCCAGTCTGCCGGGGAGGGACACAGAGGCCTTGTTA
TAAAGTCATTTACTTCCATGATACTTCTCGAAGACTGAACTTTGAGGAAGCCAAAGAAGCCT
GCAGGAGGGATGGAGGCCAGCTAGTCAGCATCGAGTCTGAAGATGAACAGAAACTGATAGAA
AAGTTCATTGAAAACCTCTTGCCATCTGATGGTGACTTCTGGATTGGGCTCAGGAGGCGTGA
GGAGAAACAAAGCAATAGCACAGCCTGCCAGGACCTTTATGCTTGGACTGATGGCAGCATAT
CACAATTTAGGAACTGGTATGTGGATGAGCCGTCCTGCGGCAGCGAGGTCTGCGTGGTCATG
TACCATCAGCCATCGGCACCCGCTGGCATCGGAGGCCCCTACATGTTCCAGTGGAATGATGA
CCGGTGCAACATGAAGAACAATTTCATTTGCAAATATTCTGATGAGAAACCAGCAGTTCCTT
CTAGAGAAGCTGAAGGTGAGGAAACAGAGCTGACAACACCTGTACTTCCAGAAGAAACACAG
GAAGAAGATGCCAAAAAAACATTTAAAGAAAGTAGAGAAGCTGCCTTGAATCTGGCCTACAT
CCTAATCCCCAGCATTCCCCTTCTCCTCCTCCTTGTGGTCACCACAGTTGTATGTTGGGTTT
GGATCTGTAGAAAAAGAAAACGGGAGCAGCCAGACCCTAGCACAAAGAAGCAACACACCATC
TGGCCCTCTCCTCACCAGGGAAACAGCCCGGACCTAGAGGTCTACAATGTCATAAGAAAACA
AAGCGAAGCTGACTTAGCTGAGACCCGGCCAGACCTGAAGAATATTTCATTCCGAGTGTGTT
CGGGAGAAGCCACTCCCGATGACATGTCTTGTGACTATGACAACATGGCTGTGAACCCATCA
GAAAGTGGGTTTGTGACTCTGGTGAGCGTGGAGAGTGGATTTGTGACCAATGACATTTATGA
GTTCTCCCCAGACCAAATGGGGAGGAGTAAGGAGTCTGGATGGGTGGAAAATGAAATATATG
GTTATTAGGACATATAAAAAACTGAAACTGACAACAATGGAAAAGAAATGATAAGCAAAATC
CTCTTATTTTCTATAAGGAAAATACACAGAAGGTCTATGAACAAGCTTAGATCAGGTCCTGT
GGATGAGCATGTGGTCCCCACGACCTCCTGTTGGACCCCCACGTTTTGGCTGTATCCTTTAT
CCCAGCCAGTCATCCAGCTCGACCTTATGAGAAGGTACCTTGCCCAGGTCTGGCACATAGTA
GAGTCTCAATAAATGTCACTTGGTTGGTTGTATCTAACTTTTAAGGGACAGAGCTTTACCTG
GCAGTGATAAAGATGGGCTGTGGAGCTTGGAAAACCACCTCTGTTTTCCTTGCTCTATACAG
CAGCACATATTATCATACAGACAGAAAATCCAGAATCTTTTCAAAGCCCACATATGGTAGCA
CAGGTTGGCCTGTGCATCGGCAATTCTCATATCTGTTTTTTTCAAAGAATAAAATCAAATAA
AGAGCAGGAAAAAAAAA

FIGURE 50

Met Arg Pro Gly Thr Ala Leu Gln Ala Val Leu Leu Ala Val Leu Leu
Val Gly Leu Arg Ala Ala Thr Gly Arg Leu Leu Ser Ala Ser Asp Leu
Asp Leu Arg Gly Gly Gln Pro Val Cys Arg Gly Gly Thr Gln Arg Pro
Cys Tyr Lys Val Ile Tyr Phe His Asp Thr Ser Arg Arg Leu Asn Phe
Glu Glu Ala Lys Glu Ala Cys Arg Arg Asp Gly Gly Gln Leu Val Ser
Ile Glu Ser Glu Asp Glu Gln Lys Leu Ile Glu Lys Phe Ile Glu Asn
Leu Leu Pro Ser Asp Gly Asp Phe Trp Ile Gly Leu Arg Arg Arg Glu
Glu Lys Gln Ser Asn Ser Thr Ala Cys Gln Asp Leu Tyr Ala Trp Thr
Asp Gly Ser Ile Ser Gln Phe Arg Asn Trp Tyr Val Asp Glu Pro Ser
Cys Gly Ser Glu Val Cys Val Val Met Tyr His Gln Pro Ser Ala Pro
Ala Gly Ile Gly Gly Pro Tyr Met Phe Gln Trp Asn Asp Asp Arg Cys
Asn Met Lys Asn Asn Phe Ile Cys Lys Tyr Ser Asp Glu Lys Pro Ala
Val Pro Ser Arg Glu Ala Glu Gly Glu Glu Thr Glu Leu Thr Thr Pro
Val Leu Pro Glu Glu Thr Gln Glu Glu Asp Ala Lys Lys Thr Phe Lys
Glu Ser Arg Glu Ala Ala Leu Asn Leu Ala Tyr Ile Leu Ile Pro Ser
Ile Pro Leu Leu Leu Leu Leu Val Val Thr Thr Val Val Cys Trp Val
Trp Ile Cys Arg Lys Arg Lys Arg Glu Gln Pro Asp Pro Ser Thr Lys
Lys Gln His Thr Ile Trp Pro Ser Pro His Gln Gly Asn Ser Pro Asp
Leu Glu Val Tyr Asn Val Ile Arg Lys Gln Ser Glu Ala Asp Leu Ala
Glu Thr Arg Pro Asp Leu Lys Asn Ile Ser Phe Arg Val Cys Ser Gly
Glu Ala Thr Pro Asp Asp Met Ser Cys Asp Tyr Asp Asn Met Ala Val
Asn Pro Ser Glu Ser Gly Phe Val Thr Leu Val Ser Val Glu Ser Gly
Phe Val Thr Asn Asp Ile Tyr Glu Phe Ser Pro Asp Gln Met Gly Arg
Ser Lys Glu Ser Gly Trp Val Glu Asn Glu Ile Tyr Gly Tyr